

Sulfoxide as a Ligand in Iron(II) Porphyrinates: S- or O-Bound?

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In order to study the bonding of sulfoxides to iron(II) porphyrinates, an equilibrium study of Fe(TPP) with tetramethylenesulfoxide (TMSO) has been performed. UV–vis spectra at different concentrations of TMSO have shown distinct character belonging to three species: four-coordinate Fe(II)(TPP), five-coordinate [Fe(II)(TPP)(TMSO)], and six-coordinate [Fe(II)(TPP)(TMSO)₂]. The isosbestic points for the low TMSO concentrations suggest that the equilibrium constant K_1 is much larger than K_2 . Analysis of spectral data by the nonlinear least-squares program SQUAD gives $K_1 = 267$ and $K_2 \approx 1$. Even though the five-coordinate species is the dominant species under the synthetic conditions, only the six-coordinate species was crystallized and characterized by an X-ray diffraction study. [Fe(TPP)(TMSO)₂] (C₅₂H₄₄Fe–N₄O₂S₂): monoclinic, $P2_1/c$, $a = 11.2580(3)$ Å, $b = 15.9262(5)$ Å, $c = 12.3930(4)$ Å, $\beta = 116.246(1)^\circ$, $V = 1992.95(10)$ Å³, $Z = 2$. X-ray crystallography demonstrates the complex is a low-spin bis-TMSO ligated species. The average Fe–N_p distance is 1.999(4) Å. The most important feature is that TMSO is coordinated to iron(II) by the sulfur donors, not oxygen. The Fe–S distance is 2.2220(3) Å.

Introduction

The amino acid methionine is a well-known ligand to iron in the ubiquitous electron-transfer proteins known as the cytochromes *c*. The other ligand is a histidine. The relative ease of losing methionine coordination has been long recognized and demonstrates the relatively small affinity of a thioether for iron.¹ Nonetheless, the coordination of the thioether sulfur atom to the physiological oxidation states of cytochrome *c*, that is, iron(II) and iron(III), is clear.

In the presence of reactive oxygen species, methionine can be oxidized to the corresponding sulfoxide.² In mitochondria (a major source of reactive oxygen species³), oxidation of methionine-80 to methionine-80 sulfoxide in the protein cytochrome *c* may occur under normal physiological conditions. Though such a transformation in nature has not been reported, the chemical modification of methionine cytochrome *c* has been studied and methionine sulfoxide cytochrome *c* has been shown fully active with cytochrome oxidase.⁴ The heme-binding characteristics of the ligand

could be changed when methionine-80 is modified to methionine-80 sulfoxide in cytochrome *c*. However, the nature of the coordination sphere in methionine-80 sulfoxide cytochrome *c* was not well established in the previously reported studies.

As an ambidentate ligand, sulfoxides can coordinate to metal ions via sulfur or oxygen.⁵ For methionine sulfoxide cytochrome *c*, an NMR⁶ study by Ivanetich et al.⁷ suggested the ligand binding is via the sulfur atom in both the ferrous and ferric forms. However, Myer concluded that modification of the methionine to methionine sulfoxide changed the sixth coordination ligand donor atom to heme iron from sulfur to the oxygen of methionine-80 sulfoxide for both iron(II) and

(5) Calligaris M.; Carugo, O. *Coord. Chem. Rev.* **1996**, *153*, 83.(6) The following abbreviations are used in this paper: NMR, nuclear magnetic resonance; AcMP8, *N*-acetylmicroperoxidase-8; TPP, dianion of *meso*-tetraphenylporphyrin; TpivotPP, dianion of $\alpha,\alpha,\alpha,\alpha$ -tetrakis(*o*-pivalamidophenyl)porphyrin; TDCP(NO₂)₂P, dianion of *meso*-tetrakis(*o*-dichlorophenyl)- β -octanitroporphyrin; PPIX, dianion of protoporphyrin IX; N_p, porphyrinato nitrogen; TMSO, tetramethylenesulfoxide; THT, tetrahydrothiophen; PMS, pentamethylene sulfide; THF, tetrahydrofuran; EtOH, ethanol; PrOH, propanol; DMSO, dimethylsulfoxide; DecMS, thioethers *n*-decylmethyl sulfide; SPh₂, diphenyl sulfide; 1-MeIm, 1-methylimidazole; 1-VinIm, 1-vinylimidazole; 1-BzylIm, 1-benzylimidazole; 4-CNPy, 4-cyanopyridine; 3-CNPy, 3-cyanopyridine; 4-MePy, 4-methylpyridine; Pc, phthalocyanine; HSAB, hard-soft-acid-base; ESR, electronic spin resonance; 18-C-6,1,4,7,10,13,16-hexaoxacyclooctadecane; 222,1,10-diaza-4,7,13,16,21,24-hexaoxabicyclo[8.8.8]hexacosane.(7) Ivanetich, K. M.; Bradshaw, J. J.; Fazakerley, G. V. *Biochem. Biophys. Res. Com.* **1976**, *72*, 433.

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Table 1. Brief Crystallographic Data and Data Collection Parameters for [Fe(TPP)(TMSO)₂]

formula	C ₅₂ H ₄₄ FeN ₄ O ₂ S ₂
fw, amu	876.88
a, Å	11.2580(3)
b, Å	15.9262(5)
c, Å	12.3930(4)
β, deg	116.246(1)
V, Å ³	1992.95(10)
space group	P2 ₁ /c
Z	2
D _c , g/cm ³	1.461
F(000)	916
μ, mm ⁻¹	0.534
crystal dimens, mm ³	0.38 × 0.33 × 0.20
radiation	Mo Kα, λ = 0.71073 Å
temp, K	100(2)
total data collected	24 711
abs corr	semiempirical from equivalents
unique data	4085 (R _{int} = 0.0200)
unique observed data [I > 2σ(I)]	3846
refinement method	full-matrix least-squares on F ²
final R indices [I > 2σ(I)]	R1 = 0.0275, wR2 = 0.0783
final R indices (all data)	R1 = 0.0293, wR2 = 0.0799

iron(III).⁸ They concluded that in the neutral-to-alkaline pH range, the low-spin spectrum is a consequence of coordination of heme iron by the sulfoxide oxygen. In recent work on AcMP8, Lushington et al.⁹ suggested that methionine-80 sulfoxide does not coordinate to iron(III) in the methionine-80 sulfoxide cytochrome *c* on the basis of the comparison of spectroscopic data for the DMSO complex of Fe(III)–AcMP8 with published data for the methionine-80 sulfoxide form. Their vacuum quantum chemical density functional theory calculations suggested that coordination of DMSO to Fe(II)–AcMP8 via oxygen is enthalpically favored over sulfur coordination; resonance Raman spectroscopic data in aqueous solution concurred with this conclusion. They claimed that the unfavorable steric environment near the heme iron in cytochrome *c* discriminates against coordination of methionine-80 sulfoxide.

Therefore, it is of fundamental importance to understand the coordination mode of sulfoxide to heme iron. Surprisingly, the interaction of sulfoxide ligand with iron(II) porphyrin has not been directly investigated. This is in distinct contrast to the situation with iron(III) where coordination of sulfoxide through oxygen has been long established.^{10,11} Reed and Scheidt have studied the sulfoxide-ligated iron(III) complex, [Fe(TPP)(TMSO)₂](ClO₄) (TMSO = tetramethylenesulfoxide), and the magnetic, Mössbauer, ESR, and structure data demonstrate that it is a high-spin O-ligated species. To our knowledge, no structural data on sulfoxide-ligated iron(II) porphyrinates have been reported so far. As part of a more general program of characterization of high-spin iron(II) porphyrinates,¹² our primary goal was

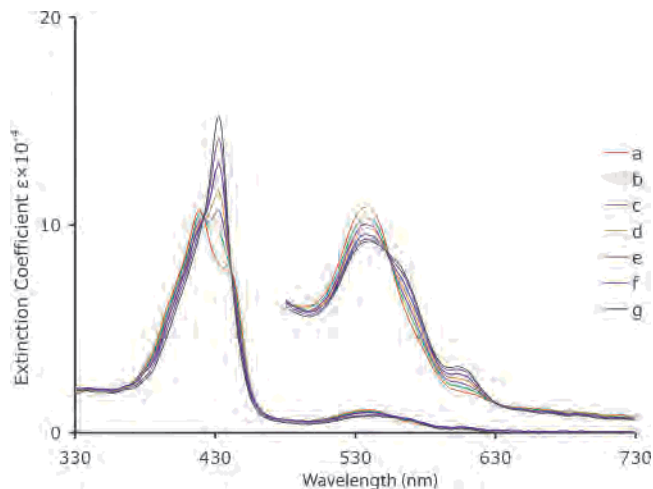


Figure 1. UV–vis spectra taken under argon in 5% EtSH/toluene solution. The concentration of [Fe(TPP)] is 0.86×10^{-4} mol/L, the TMSO concentrations in 10^{-3} mol/L (a) 1.13. (b) 2.26. (c) 3.39. (d) 5.08. (e) 7.23. (f) 9.95. (g) 14.2. The enlarged spectra from 480 to 730 nm are measured in the 10-mm UV cell.

to prepare five-coordinate TMSO-ligated species, but to our surprise, a six-coordinate low-spin S-ligated iron porphyrinate, [Fe(TPP)(TMSO)₂], was obtained. We present herein a study of the equilibria of Fe(TPP) with TMSO and the molecular structure of [Fe(TPP)(TMSO)₂].

Experimental Section

General Information. Chlorobenzene was washed with concentrated sulfuric acid, then with water until the aqueous layer was neutral, dried with MgSO₄, and distilled twice over P₂O₅ under argon. Hexanes were distilled over sodium benzophenone. Ethanethiol (Aldrich) was used as received. All other chemicals were used as received from Aldrich or Fisher. *meso*-Tetraphenylporphyrin (H₂TPP) was prepared according to Adler et al.¹³ [Fe(TPP)Cl] was prepared according to a modified Adler preparation.^{14,15} [Fe(TPP)]₂O was prepared from [Fe(TPP)Cl].¹⁵ All manipulations were performed under argon.

Synthesis of [Fe(TPP)(TMSO)₂]. [Fe(II)(TPP)] was prepared by reduction of [Fe(TPP)]₂O (32 mg, 0.024 mmol) with ethanethiol (~1 mL) in ~6 mL of chlorobenzene. The chlorobenzene solution was stirred for 3 days, then transferred into a Schlenk flask containing tetramethylenesulfoxide (0.5 mL, 5.6 mmol) by cannula, and stirred for another hour. X-ray quality crystals were obtained in sealed 8 mm diameter glass tubes by liquid diffusion using hexanes as the nonsolvent.

Equilibrium Constant Determination. UV–vis spectra were recorded on a Perkin-Elmer Lambda 19 UV–vis–near-IR spectrometer in a specially designed combined 1- and 10-mm inert atmosphere cell. A solution of the four-coordinate iron(II) porphyrin was prepared as above. UV–vis spectra of Fe(TPP) in different concentrations of TMSO were measured. For the low-concentration region, TMSO was titrated into [Fe(II)(TPP)] in a 5% EtSH/toluene

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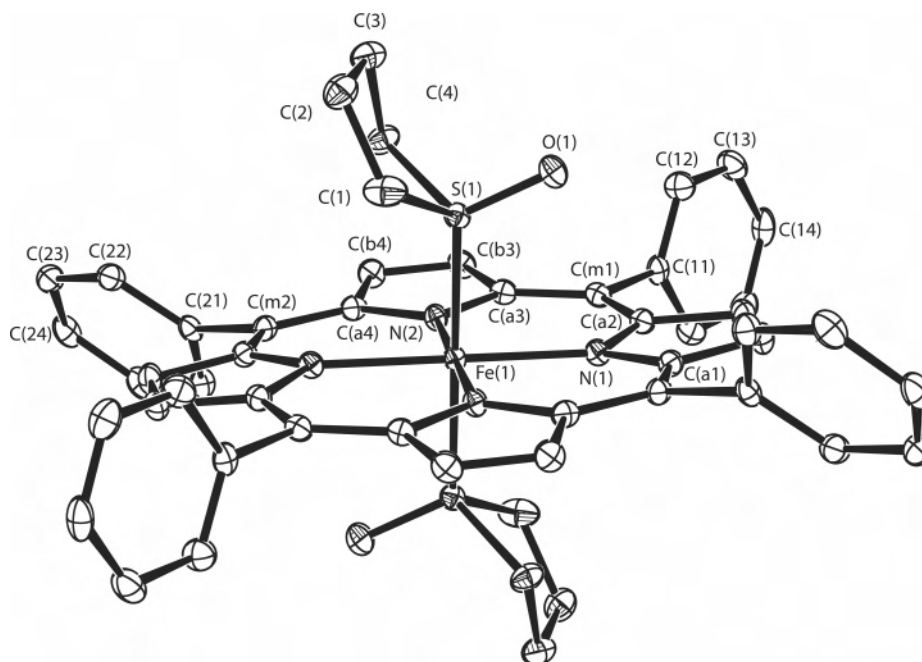


Figure 2. ORTEP diagram of $[\text{Fe}(\text{TPP})(\text{TMSO})_2]$. The hydrogen atoms of the porphyrin ligand have been omitted for clarity; 50% probability ellipsoids are depicted.

solution; the EtSH was added to prevent oxidation of the iron.¹⁶ For the high-concentration region, the $[\text{Fe}(\text{II})(\text{TPP})]$ solution was added into the concentrated TMSO solution. In order to obtain spectra with appropriate intensity for both the Soret and Q bands, several different concentrations of iron porphyrin were used. The concentrations of TMSO for UV–vis measurements range from 1.13×10^{-3} to 10.7 mol/L. The equilibrium constants for these equilibria were calculated with the nonlinear least-squares program SQUAD.¹⁷ SQUAD calculates best values for the equilibrium constants by employing multiple-wavelength absorption data for varying concentrations of the reactants. Absorbance data, at 10-nm increments spanning the visible region, were input into SQUAD.

X-ray Structure Determination. Single-crystal experiments were carried out on a Bruker Apex system with graphite-monochromated Mo $K\alpha$ radiation ($\lambda = 0.71073 \text{ \AA}$). A red crystal with the dimensions $0.38 \times 0.33 \times 0.20 \text{ mm}^3$ was used for the structure determination. The crystalline sample was placed in inert oil, mounted on a glass pin, and transferred to the cold gas stream of the diffractometer. Crystal data were collected at 100 K. The structure was solved by direct methods using SHELXS-97¹⁸ and refined against F^2 using SHELXL-97;^{19,20} subsequent difference Fourier syntheses led to the location of all the remaining non-hydrogen atoms. For the structure refinement, all data were used

including negative intensities. The structure was refined in space group $P2_1/c$. The iron atom is located at a crystallographic inversion center, and the asymmetric unit contains half an iron porphyrin and one TMSO ligand. All non-hydrogen atoms were refined anisotropically. Hydrogen atoms were added with the standard SHELXL-97 idealization methods. The program SADABS²¹ was applied for the absorption correction. Brief crystal data are listed in Table 1. Complete crystallographic details, atomic coordinates, anisotropic thermal parameters, and hydrogen atom coordinates are given in the Supporting Information.

Results

Figure 1 shows the UV–vis spectra of $\text{Fe}(\text{TPP})$ with TMSO concentrations ranging from 0 to 1.49×10^{-1} mol/L; the highest concentrations of TMSO are not illustrated. Before the addition of TMSO, the spectrum of $\text{Fe}(\text{II})(\text{TPP})$ shows two Soret bands at 419 and 442 nm and a relative weaker band at 537 nm in the Q -band region (500–700 nm). During the titration, the electronic spectra show that the Soret band becomes a single sharp band at 432 nm with increased intensity. The band at 537 nm decreases, and new bands show up at 560 and 603 nm. Clear isosbestic points are observed in both the Soret and Q -band regions. When the concentration of TMSO is over about 0.45 mol/L, the Soret band begins to decrease and finally shifts to 429 nm with a relatively weaker intensity in 10.7 mol/L of TMSO (95%). The bands at 560 and 603 nm decrease, and the band at 530 nm increases by the increasing of TMSO concentration.

Crystals were obtained from solutions where the TMSO-to- $\text{Fe}(\text{TPP})$ concentration ratio was ~ 220 . X-ray crystallography shows that the isolated complex is a six-coordinate

(16) Even under these dilute solution conditions, we see no spectral changes resulting from the presence of EtSH and we conclude that EtSH does not coordinate to iron. The relatively long times of the titrations do require EtSH.

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(20) $R1 = \sum ||F_o| - |F_c|| / \sum |F_o|$ and $wR2 = \{[\sum (F_o^2 - F_c^2)^2] / [\sum w(F_o^4)]\}^{1/2}$. The conventional R factors $R1$ are based on F , with F set to 0 for negative F^2 . The criterion of $F^2 > 2\sigma(F^2)$ was used only for calculating $R1$. R factors based on $F^2(wR2)$ are statistically about twice as large as those based on F , and R factors based on all data will be even larger.

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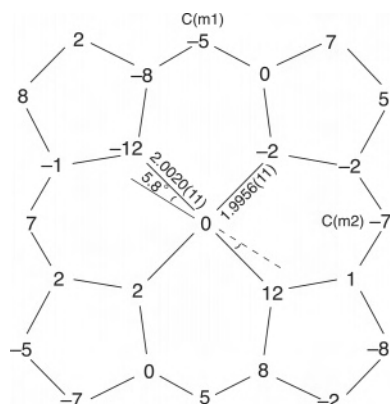


Figure 3. Diagram illustrating the core conformation and iron displacement of the title complex. The displacements of the iron and the atoms of the porphyrin core from the mean plane defined by the 24 porphyrin atoms are given. The position of the S–O bond with respect to directions defined by the Fe–N_p directions is shown: solid lines are above the plane, dashed lines are below the plane.

Table 2. Selected Bond Lengths (Å) and Angles (deg) for for [Fe(TPP)(TMSO)₂]

Fe(1)–N(1)	2.0020(11)	Fe(1)–N(2)	1.9956(11)
Fe(1)–S(1)	2.2220(3)	S(1)–O(1)	1.4794(11)
S(1)–C(4)	1.7981(14)	S(1)–C(1)	1.8076(15)
N(2)–Fe(1)–N(1)	90.55(5)	N(1)–Fe(1)–S(1)	87.62(3)
N(2)–Fe(1)–S(1)	90.63(3)	O(1)–S(1)–Fe(1)	118.91(5)
O(1)–S(1)–C(4)	106.54(7)	O(1)–S(1)–C(1)	106.57(7)
C(4)–S(1)–C(1)	91.76(7)	C(2)–C(1)–S(1)	107.22(10)
C(4)–S(1)–Fe(1)	115.18(5)	C(1)–S(1)–Fe(1)	114.27(5)

species. An ORTEP diagram is provided in Figure 2, and selected bond distances and angles are listed in Table 2. The porphyrin molecule has inversion symmetry, the iron(II) atom is at an inversion center and coordinated to four pyrrole nitrogens and two sulfur atoms of TMSO. The average Fe–N_p distance is 1.999(4) Å, and the Fe–S distance is 2.2220(3) Å. Coordination at the sulfur atom is pyramidal, with an O–S–C bond angle of 106.54(7)°, a C–S–C angle of 91.76(7)°, a Fe–S–O angle of 118.91(5)°, and an average Fe–S–C angle of 114.7(6)°. The S–O distance is 1.4794(11) Å. A complete listing of bond distances and angles is given in the Supporting Information.

The displacement of each atom of the porphyrin core from the 24-atom mean plane is shown in Figure 3. Also included on the diagrams of Figure 3 are the individual Fe–N_p bond lengths. An analogous diagram showing atomic displacements from the mean plane of the four pyrrole nitrogen atoms is given in Figure S1 of the Supporting Information.

Discussion

Spectral Studies. There are three species involved in the reaction of TMSO with Fe(II)(TPP): four-coordinate Fe(II)(TPP), five-coordinate [Fe(II)(TPP)(TMSO)], and six-coordinate [Fe(II)(TPP)(TMSO)₂].

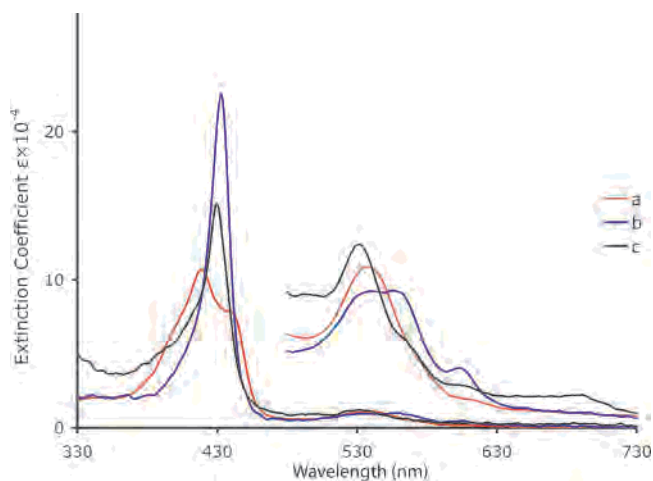
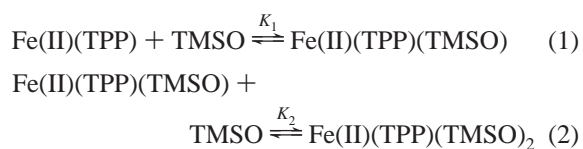


Figure 4. UV–vis spectra taken under argon. (a) Fe(II)(TPP) (8.6×10^{-5} mol/L). (b) Fe(II)(TPP) (3.8×10^{-5} mol/L) in 4.3×10^{-2} mol/L TMSO solution. (c) Fe(II)(TPP) (3.0×10^{-5} mol/L) in 10.7 mol/L TMSO solution. The enlarged spectra from 480 to 730 nm are measured in the 10-mm UV cell.

Because the iron(II) porphyrinates with different coordination numbers have substantially different absorption spectra,²² UV–vis spectroscopy can be used to monitor the reaction and determine the equilibrium constants. As shown in Figures 1 and 4, both the Soret band and Q bands show remarkable changes during the titration. The spectrum with TMSO concentration at 4.3×10^{-2} mol/L shows a sharp Soret band at 432 nm and three bands in the Q-band region: two broad bands with similar intensities at around 540 and 555 nm and one at 603 nm with about half the intensity of the other two bands. This spectral pattern, including intensity, is similar to that observed for imidazole-ligated five-coordinate high-spin iron(II) species.²² We can thus conclude that the dominant species in this solution is a five-coordinate high-spin species. Analysis by the SQUAD program suggests [Fe(TPP)(TMSO)] is 89% of the total iron species at this concentration. The spectrum in 10.7 mol/L TMSO is substantially different. The single Soret band is shown at a similar position with lower intensity. The Q bands show distinct differences in extinction coefficients: the major peak at 530 nm is about twice as intense as the shoulder peak at 560 nm. These features are the same as features observed in the six-coordinate nitrogen-based species,²² which suggests the dominant species in this solution is six-coordinate low-spin [Fe(TPP)(TMSO)₂]. Analysis by the SQUAD program confirms [Fe(TPP)(TMSO)₂] is 91% of the total iron species at this concentration. Values of the extinction coefficients are calculated by the SQUAD program: λ_{max} (log ϵ): 432–(5.38), 540(4.03), 560 (4.03), 603(3.61) for [Fe(TPP)(TMSO)]; 429(5.16), 530 (4.13), 560(3.96) for [Fe(TPP)(TMSO)₂].

The spectral changes in Figure 1 suggests that they relate to reaction 1. The appearance of isosbestic points indicates only one major species formed at these concentrations, a five-coordinate species. It also suggests that the equilibrium

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Table 3. Selected Bond Distances (Å) and Angles (deg) for [Fe(TPP)(TMSO)₂] and Related Species

complex	Fe–N _p ^a	Fe–S _{ax}	Fe–O _{ax}	S–O	spin state ^b	ref
[Fe(TPP)(TMSO) ₂]	1.999(4)	2.2220(3)		1.4794(11)	LS	tw
[Fe(TPP)(TMSO) ₂]ClO ₄	2.046(6)		2.078(13)	1.522(9)	HS	10
Fe(OEP)(DMSO) ₂ PF ₆	2.035(9)		2.082(5)	1.539(6)	HS	11
[Fe(TPP)(THT) ₂]	1.996(6)	2.336(3)			LS	25
[Fe(TPP)(THT) ₂]ClO ₄	1.985(3)	2.357(27)			LS	25
[Fe(TPP)(PMS) ₂]ClO ₄	1.982(6)	2.341(1)			LS	25
[Fe(II)(TPP)(THF) ₂]	2.057(4)		2.351(3)		HS	36
[Fe(II)(F ₈ TPP)(THF) ₂]	2.068(9)		2.314(3)		HS	37
[Fe(TTP)(THF) ₂]	2.067(7)		2.3208(8)		HS	38
[Fe(TDCP(NO ₂) ₈ P)(EtOH) ₂]	2.074(6)		2.153(11)		HS	39
[Fe(TDCP(NO ₂) ₈ P)(PrOH) ₂]	2.092(5)		2.152(2)		HS	39

^a Average Fe–N_p distance. ^b LS, low spin; HS, high spin.

constant K_1 is much larger than K_2 . The analysis of the spectral data by the SQUAD program supports this. The result gives formation constants $\log K_1 = 2.43 \pm 0.06$ and $\log \beta_2 = \log(K_1K_2) = 2.41 \pm 0.16$. This gives $K_1 = 267$ and $K_2 = 0.95$. A value of $K_2 \approx 2300$ has been reported from a photolysis experiment.²³ It is likely that the equilibrium constant (in pure TMSO) is actually that of K_1 .

All the above suggests three species in solution are dominant at different concentrations of TMSO. Six-coordinate [Fe(TPP)(TMSO)₂] is the dominant species in solution at very high concentrations of TMSO, while the five-coordinate species is the dominant species at relatively low concentrations. In order to prepare a five-coordinate species, the synthesis was performed at 0.75 mol/L of TMSO. At that concentration, the dominant species in solution is five-coordinate [Fe(TPP)(TMSO)], but the six-coordinate species was crystallized and verified by X-ray crystallography. Its crystallization is probably due to solubility issues even though it is not the major species in solution. Finally, it should be noted that our spectroscopic data do not define the coordination modes (O vs S) nor that in a mixed ligand complex such as a nitrogen donor/sulfoxide that iron(II) would definitely be coordinated by the sulfur.

Molecular Structure. In the crystal structure, the average Fe–N_p distance in [Fe(TPP)(TMSO)₂] is 1.999(4) Å. This distance is that expected for a low-spin iron(II) species.²⁴ Low-spin iron(II) examples for comparison include a thioether-ligated species, [Fe(TPP)(THT)₂] (1.996(6) Å);²⁵ several imidazole-ligated species, [Fe(TPP)(1-VinIm)₂] (2.001(2) Å), [Fe(TPP)(1-BzylIm)₂] (1.993(9) Å), and [Fe(TPP)(1-MeIm)₂] (1.997(6) Å);²⁶ and several pyridine-ligated species, [Fe(TMP)(4-CNPy)₂] (1.992(1) Å), [Fe(TMP)(3-CNPy)₂] (1.996(0) Å), and [Fe(TMP)(4-MePy)₂] 1.988(0) Å.²⁷

The low-spin state of [Fe(TPP)(TMSO)₂] is in distinct contrast to the bis(sulfoxide) complexes of iron(III) porphyrinates that are all known to be high spin.^{10,11} The significant difference between the iron(II) and iron(III) porphyrinate systems is the ligation mode of the sulfoxide ligands: O-bound in iron(III) and S-bound in iron(II). Qualitatively, such a difference in axial coordination mode could be predicted by Pearson's HSAB principles.²⁸ The harder iron(III) is expected to bind the hard oxygen donor atom of the sulfoxide, consistent with observation. Iron(II), on the other hand, is considered a borderline acid and either O- or S-ligated sulfoxide species might be anticipated. Indeed, both

O-^{29,30} and S-bound^{31,32} sulfoxide complexes of iron(II) are known. However, the "hardness" and "softness" of a metal ion can be dramatically modified by the nature of the other ligands, depending on their σ -donating or π -accepting properties.^{33,34} For the case of porphyrin derivatives, the highly delocalized π system will tend to make iron(II) softer and favor sulfur ligation as observed.

Most oxygen-ligated iron(II) porphyrinates are species with THF or alcohol as the axial ligands.^{35–39} Contrasting with the aforementioned low-spin S-ligated iron(II) porphyrinates, their central irons are in the high-spin state with an expanded porphyrin core, as shown in Table 3. As can be seen in the Figure 3, the porphyrin core is very planar. The S–O bond is close to eclipsing the N(1)N(1') direction, (N(1') is a symmetry related pyrrole nitrogen of N(1)). The angle between the projection of the S–O bond, and the Fe–N_p vector is 5.8°. The orientation is also illustrated in Figure 3.

The six-coordinate, S-ligated state of [Fe(TPP)(TMSO)₂] is clearly seen in Figure 2. Scheidt and Reed had noted earlier that the Fe–S distances in a wide variety of iron porphyrinate derivatives was insensitive to a number of factors that could

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be expected to lead to differing Fe–S distances: all observed distances are ~ 2.35 Å. The relatively short Fe–S distance of 2.2220(3) Å in [Fe(TPP)(TMSO)₂] was therefore rather unexpected and can be compared to 2.338(1), 2.334(1) Å in [Fe(TPP)(THT)₂],²⁵ 2.367(3) Å in K(18-C-6)[Fe(TpivPP)(SC₆HF₄)(O₂)], 2.365(2) Å in Na(18-C-6)[Fe(TpivPP)(SC₆HF₄)(O₂)],⁴⁰ 2.324(2) Å in Na(222)[Fe(TpivPP)(SC₂H₅)],⁴¹ and 2.380(2) Å in K(222)[Fe(TpivPP)(PMS)(NO₂)].⁴² It is thus seen that the Fe–S distance in [Fe(TPP)(TMSO)₂] is more than 0.1 Å shorter than all previously observed Fe^{II}–S distances. A similar situation has been seen in ruthenium(II) porphyrinates, although the difference between Ru–S distances in the S-ligated sulfoxide and thioether complexes is about 0.05 Å. The observed distances are Ru–S = 2.319(1) Å in [Ru(II)(OEP)(DMSO)₂],⁴³ and 2.376(1) and 2.371(1) Å in [Ru(OEP)(DecMS)₂] and [Ru(OEP)(SPh₂)₂],⁴⁴ respectively.

The shortening of the Fe–S bond in the S-bound sulfoxides relative to analogous thioether derivatives can be associated with either a strengthening of Fe–S $d_{\pi}-p_{\pi}$ back-bonding, enhanced σ -bonding, or possible steric effects. Steric effects appear to have no influence as nonbonded distances between the TMSO ligand or the tetrahydrothiophene ligand and the porphyrin in the respective six-coordinate complexes are similar. Increased π -back-bonding has been proposed for Ru(II)–DMSO nonporphyrin complexes;^{5,45,46} such π bonding is associated with a lengthening of the S–O bond relative to the value in uncomplexed sulfoxides. However, in [Fe(TPP)(TMSO)₂], the S–O distance (1.4794(11) Å) is close to that in free TMSO (1.484(3) Å).⁴⁷ Thus,

π -back-bonding does not appear to be making a significant contribution to the shortness of the Fe–S bond in [Fe(TPP)(TMSO)₂]. Enhanced σ -bonding of the TMSO is the most possible reason for the shortening of the Fe–S bond. If there is enhanced σ -bonding in TMSO complexes, the TMSO will be expected to be more basic than tetrahydrothiophene which has been observed.⁴⁸ This enhanced σ -bonding has also been suggested by Lushington et al.⁹ in a DFT calculation.

Summary

A study of the equilibria of Fe(TPP) with TMSO has been performed. Equilibrium constants were obtained as $K_1 = 267$ and $K_2 = 0.95$. At low concentration, the spectra show isosbestic points during the titration. Three species in solution are dominant at different TMSO concentrations. Under the synthetic conditions, the dominant species in solution is five-coordinate species. But probably due to solubility issues, the six-coordinate species, [Fe(II)(TPP)(TMSO)₂], was crystallized. X-ray crystallography shows that is a sulfur-ligated low-spin species, not like the oxygen-ligated high-spin [Fe(III)(TPP)(TMSO)₂]ClO₄. The average Fe–N_p distance is 1.999(4) Å, and the Fe–S distance is 2.2220(3) Å.

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Supporting Information Available: Figure S1 displays displacements from the N₄ plane; Tables S1–S6 give complete crystallographic details, atomic coordinates, bond distances and angles, anisotropic temperature factors, and fixed hydrogen atom positions; and X-ray crystallographic data in CIF format. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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